

Europäisches Patentamt

Eur pean Patent Office

Office européen des brevets



(11) EP 0 882 448 A1

(12)

EUROPEAN PATENT APPLICATION

(43) Date of publication: 09.12.1998 Bulletin 1998/50

(51) Int. Cl.⁶: **A61K 9/50**

(21) Application number: 97830211.5

(22) Date of filing: 05.05.1997

(84) Designated Contracting States: DE FR GB IT NL

(71) Applicant: DIDECO S.p.A.
I-41037 Mirandola (Province of Modena) (IT)

(72) Inventors:

- Magnani Mauro, 61029 Urbine (IT)
- Panzani Ivo, 41037 Mirandola (IT)

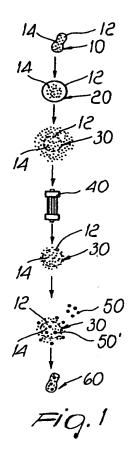
- Bigi Leonardo, 41037 Mirandola (IT)
 Zanella Andrea, 41037 Mirandola (IT)
- (74) Representative:

 Modiano, Guido, Dr.-Ing. et al

 Modiano & Associati SpA

 Via Meravigli, 16

 20123 Milano (IT)
- (54) Method of encapsulating biologically active agents within erythrocytes and apparatus therefor
- (57) A method and apparatus for the encapsulation of biologically active agents in red blood cells (RBCs) is disclosed. The method is characterized in that lysed and partially lysed RBCs are concentrated prior to the introduction of the biologically active agent. The resultant carrier RBCs are stable for an extended period of time under physiological conditions and can be used for the systemic or targeted delivery of therapeutically effective amounts of one or more biologically active agents to a patient.



Description

Field of the Invention

The present invention relates to a method for the incorporation of biologically active agents into red blood cells.

Background of the Invention

Much effort has been recently focused on the development of procedures for the targeted delivery of pharmaceutical agents to specific sites in a patient. It is known that drug efficacy can be increased when the appropriate target site is efficiently reached, and drug toxicity can be reduced when the total amount of drug administered is minimized. The interest in drug delivery systems applies both to conventional agents, many of which are relatively simple organic molecules, as well as to more complex agents such as oligopeptides, prot ins and nucleic acids, etc. One area of recent interest relates to the use of red blood cells (hereafter erythrocyt s and RBCs) to deliver therapeutic dosages of drugs to a target site in a patient. Erythrocytes can be "loaded" with biologically active agents by a process in which the cell membranes of the erythrocytes are lysed and one or more agents is added to the erythrocytes, followed by resealing of the cell membranes. Such "loaded" or "carrier" erythrocytes offer a number of advantages as drug delivery and targeting systems because they are biodegradable, can be maintained in the circulatory system for long periods of time, and can be targeted to selected cells such as, for example, macrophages. However, although the use of erythrocytes as drug delivery systems has been investigated by many, the method has not yet developed to the point where it can be applied routinely in a clinical setting.

Processes for the preparation of a suspension of loaded cells in a physiological solution are disclosed in German Patent No. 23 26 244 and in German published Patent Application Nos. (OS) 23 26 161 and 24 95 119, in which the erythrocyte cell membranes are lysed by either osmotic pressure or an electrical field, respectively.

U.S. Patent No. 4,224,313, (Zimmermann et al.), discloses a method for preparing a mass of loaded cells suspended in a solution by increasing the permeability of the cell membranes via externally induced osmotic pressure or an electric field, or both. The material to be loaded includes a pharmaceutical agent having a capability, when incorporated in a cell, of prematurely destroying cell membranes, and a stabilizing agent capable of inhibiting the reaction of the pharmaceutical agent with the cell membranes.

U.S. Patent No. 4,269,826, (Zimmermann et al.), discloses a method for incorporating magnetic substances within loaded cells that can be localized to a specific part of a living body by application of an exter-

nal magnetic field.

U.S. Patent No. 4,289,756, (Zimmermann et al.), discloses a method for preferentially accumulating loaded cells within the spleen and liver of a living body.

U.S. Patent No. 4,478,824, (Franco et al.), discloses a method for incorporating substances into erythrocytes by changing the internal osmotic pressure of RBCs by the action of chemical agents, such as DMSO and glycerol, that can pass through the cell membrane and enter cells by diffusion.

U.S. Patent No. 4,652,449, (Ropars et al.), discloses a method and apparatus for incorporating materials into erythrocytes using osmotic pressure. The method and apparatus have been used and tested only with large volumes of blood, which limits many applications to the use of homologous blood, i.e., blood pooled from many individual sources. U.S. Patent No. 4,931,276, (Franco et al.), discloses a method for the encapsulation of non-ionic agents within erythrocytes. The method is of limited effectiveness where the desired agent to be incorporated is not anionic, or is anionic or polyanionic but is not present in the near-isotonic aqueous medium in sufficient concentration to cause needed increases in cell permeability without cell destruction.

Heubsch et al., J. Cell. Physiol., 122:266-272 (1985), demonstrate that cytoskeletal detachment occurs in osmotically swollen cells and that with extreme changes in size, the bilayer membrane is released from a deformational constraint which is present under normal conditions.

Reviews of methods of incorporating substances into cells are presented by Franco et al. in Life Science 32:2763-2768 (1983), Am. J. Hematol. 17:393-400 (1984), and J. Cell. Physiol. 129:221-229 (1986).

Objects and Summary of the Invention

Despite the extensive work in the field of the present invention, there is still no method known in the prior art which provides for the introduction of one or more biologically active agents into RBCs at acceptable physiological concentrations without the dilution or loss of the endogenous contents of the erythrocytes. It is a primary object of the present invention to provide a method which fulfills this need. That is, the method of the invention provides for the introduction of one or more biologically active agents (i.e., drugs) into erythrocytes at desirable physiological concentrations without the dilution or loss of a substantial portion of the cellular contents, which undesirably decreases the integrity, viability and longevity of carrier RBCs.

More particularly, it is an object of the present invention to provide such a method using a small volume of blood so that autologous carrier RBCs may be prepared for individual patients, and without the need for cooling of the blood at any stage of the process.

It is a further object of the invention to provide car-

Serve Server - -

25

40

45

50

rier (i.e., drug-loaded) erythrocytes that possess the ability to persist, and to release biologically active agents, under mammalian physiological conditions for a longer period of time than previously described carrier RBCs. Carrier RBCs made according to the present invention release their encapsulated biologically active agent(s) by diffusion out of the RBCs or by active transport across the erythrocyte cell membrane.

It is a further object of the present invention to provide an apparatus by which the method of the invention can efficiently and reproducibly carried out. More particularly, it is an object of the present invention to provide an apparatus by which autologous carrier RBCs may be prepared from a small volume of blood from an individual patient in a matter of hours.

Thus, the invention relates to the incorporation of a wide variety of therapeutically useful biologically active agents into mammalian erythrocytes which could not previously be accomplished without unacceptable dilution of the endogenous erythrocyte contents and/or the agent(s) to be incorporated within the erythrocytes. More particularly, the method of the present invention makes possible the introduction or incorporation of various agents, including polypeptides, into erythrocytes. These and other biologically active agents may be encapsulated, either alone or in combination with other such agents, within erythrocytes that thus become carrier erythrocytes for the agent(s). The carrier erythrocytes are used to achieve a desired slow continuous delivery of the encapsulated biologically active agent(s) when the carrier erythrocytes are later introduced into a mammal. Moreover, the invention provides for the rapid preparation of such carrier erythrocytes from relatively small samples of blood and thus allows for the preparation of autologous carrier erythrocytes from an individual patient. The method of the invention has particular utility when used with an apparatus as described herein.

The above objects are achieved, in accordance with the present invention, in a method of introducing at least one desired agent into mammalian red blood cells (RBCs), which comprises concentrating lysed or partially lysed erythrocyte material before adding one or more biologically active agents to be encapsulated and which significantly improves the encapsulation yield. Concentration of the lysed or partially lysed erythrocyte material can be achieved by a variety of means, including hemofiltration, which uses a high flux hemodialyzer such as a hollow fiber dialyzer with a high ultrafiltration rate. In a preferred embodiment, a disposable hemofilter cartridge is used as the concentrating means in the method of the invention.

Brief Description of the Drawings

FIG. 1 schematically shows one preferred embodiment of the method of the invention.

FIG. 2 is a schematic representation of one embodiment of an apparatus useful for carrying out the method of the present invention.

Detailed Description of the Invention

The method of the present invention provides for the introduction of at least one desired agent into mammalian red blood cells. The method involves concentrating lysed or partially lysed erythrocyte material before adding one or more biologically active agents to b encapsulated and has been found to significantly improve the encapsulation yield. As used herein, "encapsulation yield" refers to the fraction of a biologically active agent that is encapsulated within carrier RBCs, and is usually expressed as the percentage of encapsulated agent relative to the total amount of agent added during the encapsulation process. The erythrocyte material can be concentrated by a variety of means, including hemofiltration, which uses a high flux hemodialyzer such as a hollow fiber dialyzer with a high ultrafiltration rate. In a preferred embodiment, a disposable hemofilter cartridge is used as the concentrating means in the method of the invention.

As used herein, "carrier RBCs" refers to erythrocytes which encapsulate one or more biologically active agents. In some instances, carrier RBCs containing an added biologically active agent are described herein with respect to the specific agent. For example, "d-21P carrier RBCs" refers to erythrocytes that have be n lysed, loaded with the agent dexamethasone-21-phosphate (d-21P), and release dexamethasone. Carrier RBCs for a variety of such agents, either alone or in combination, can be prepared by the method of th invention.

As used herein, biologically active agents refers to materials capable of being encapsulated using the methods of the present invention. These include, but are not limited to, peptides, oligopeptides, polypeptides, proteins, hormones, corticosteroids, glucocorticoids, non-steroidal anti-inflammatory agents, glutathion, cytokines, toxins, oligonucleotides and other nucleic acids, and nucleoside analogs that are well known as useful therapeutic agents. These include 6-mercaptopurine (6-MP) or azathiopurine and fludarabine phosphate, which are commonly used immunosuppressants and inhibitors of malignant cell growth, and phosphorylated azidothymidine (AZT), dideoxycytosine (ddC) and dideoxyinosine (ddI), which are useful as anti-viral agents, particularly in the treatment of AIDS. Carrier RBCs made by the method of the present invention have the desirable attribute of enhanced integrity, viability and longevity under physiological conditions.

The following abbreviations are used throughout:

BS³ bissulfosuccinimidilsuberate dexamethasone-21-phosphate

PIGPA Phosphate-Inosine-Glucose-Pyruvate-Ade-

nine

25

RBCs red blood cell(s), erythrocytes

CPD Sodium citrate dihydrate-Citric acid monohy-

drate-Sodium phosphate monobasic bihy-

drate-Glucose

PBS⁽¹⁾ phosphate buffered saline

In one embodiment, the method of the present invention comprises collecting erythrocytes from an sindividual patient, concentrating the erythrocytes, washing the erythrocytes with a first solution of a physiological saline, washing the erythrocytes with a second. hypotonic solution consisting of 6 volumes of a physiological saline solution and 4 volumes of distilled sterile H₂O, diluting the erythrocytes with a solution consisting of 6 volumes of phosphate buffered saline (PBS) and 8 volumes of distilled sterile H2O, concentrating of the erythrocytes in a hemofilter cartridge, adding the biologically active agent or agents to be encapsulated, resealing the erythrocytes by addition of a Phosphate-Inosine-Glucose-Pyruvate-Adenine (PIGPA) solution, incubating the resulting erythrocytes at about 37°C for about 30 minutes, and washing the erythrocytes with a physiological saline solution. The PIGPA solution preferably comprises about 33 mM NaH₂PO₄; about 1.606 M KCl; about 0.194 M NaCl; about 0.1 M inosine; about 5 mM adenine; about 20 mM ATP; about 0.1 M glucose; about 0.1 M pyruvate; and about 4mM MgCl₂. In a preferred embodiment, about 5 ml of the PIGPA solution is used for resealing a volume of about 70-90 ml of lysed erythrocytes.

In order to prevent the erythrocytes from coagulating during the process, they may be optionally contacted with an anticoagulant during or subsequent to the collection step. One preferred anticoagulant solution, referred to herein as CPD solution comprises 26.3 g sodium citrate dihydrate, 3.3 g citric acid monohydrate, 2.5 g sodium phosphate monobasic bihydrate, and 23.2 g glucose in 1 liter of sterile water.

More particularly, the method of the present invention involves separating erythrocytes from other blood cells and components, suspending the erythrocytes in a first hypotonic solution at a concentration of about 0.04 to about 0.05 ml of erythrocytes for each ml of the first solution (the volume and osmolality of the first solution being known or determined), lysing the erythrocytes by suspending them in a second hypotonic solution (having an osmolality that is about 30 to about 60 mOsm less than that of the first solution) to obtain a lysate, adjusting the concentration of the lysate so that its volume is about 0.1 to about 0.067 that of the volume of the first solution, adding a biologically active agent to be encapsulated to the lysate, and resealing the lysed RBCs by adding a concentrated saline solution to the lysate to adjust the osmolality of the lysate to a range of about 1.9 to about 2.1 times that of the first solution.

(1)Physiological saline (NaCl 0.9% w/v) may be substituted for PBS.

A preferred embodiment of the method of the invention is shown schematically in FIG. 1. In FIG. 1, erythrocytes 10 are represented as a membrane 12 containing numerous small filled circles. The small filled circles are intended to represent biological molecules 14 endogenous to unlysed RBCs. After the first hypotonic dilution, it is seen that the erythrocytes 10 become swollen (schematically represented as 20). The swollen erythrocytes 20 are exposed to a second hypotonic dilution, which causes them to become fully or partially lysed. The lysed or partially lysed erythrocytes 30 both enclose, and are surrounded by, biological molecules 14 endogenous to unlysed RBCs. In the next step, the lysed or partially lysed erythrocytes 30 are then concentrated in a hemofilter 40. The resulting lysed or partially lysed erythrocytes 30 are contacted with biologically active agents 50 represented by the large filled circles. In so doing, some of the biologically active agents become encapsulated within the erythrocytes. The encapsulated biologically active agents are schematically represented as 50. The erythrocytes, now containing biologically active agents, are then exposed to a concentrated saline solution. The exposure causes the cell membranes 12 to reseal, thereby capturing and encapsulating biologically active agents 50 within the cell. The resulting, so-called carrier RBC or carrier erythrocyte 60 is then washed.

One embodiment of apparatus for carrying out the method described above is shown schematically in FIG. 2. In FIG. 2, a bag 100 preferably containing at least 50 ml of blood (collected from a patient) is connected to a blood feed line 102. The blood is transferred, by a pump 104 to a separation bowl 106, preferably rotating at about 5600 rpm. Erythrocytes contained within the bowl 106 are washed by a physiological saline solution (NaCl 0.6%) provided by a physiological saline feed line 108. The physiological saline is preferably supplied at a rate of about 150 ml/min. The washing procedure separates plasma and the other blood cells which are transferred to a waste collection bag 110. After washing, the erythrocytes are pumped from the separation bowl 106 to a transfer bag 112. About 400 ml of a first hypotonic solution (6 volumes physiological saline and 4.5 volumes distilled water) are added to the transfer bag 112 via a first hypotonic solution feed line 114 and mixed with the erythrocytes. The resulting suspension is then maintained at room temperature for about 5-10 minutes under gentle agitation. At this point the erythrocytes have become swollen (reference numeral 20 of FIG. 1), but they have not yet been lysed. A second hypotonic solution is then introduced into the transfer bag 112 via a second hypotonic solution feed line 116. The second hypotonic solution is preferably about 200 ml of a solution having 6 volumes physiological saline and 8 volumes distilled water. It is at this stage that hemolysis of the erythrocytes occurs. After about 15 minutes at room temperature, the resulting lysate is pumped through the hemofilter 118. During the filtration process, clamps

25

30

35

120, 122, 124 and 126 are open, and clamps 128, 130, 132, 134, 136, 138 and 140 are closed. To enhance the filtration process, an optional vacuum pump 142, drawing a vacuum of about -100 to about - 150 mmHg, may be employed. Hemolysate is collected in an ultrafiltrate reservoir 144 until the volume of the filtrate is in the range of about 170-190 ml. Once the desired volume has been achieved, the concentrated erythrocytes are collected in the transfer bag 112. This is accomplished by closing clamp 124 and opening clamp 128. The biologically active substance is then introduced into the transfer bag 112, at which point it contacts and becomes encapsulated by the concentrated, lysed erythrocytes. After about 10-15 minutes of gentle agitation, 5 ml of PIGPA resealing solution is added to the transfer bag 112, and the bag is placed into a heater 146 for about 30 min at about 37 °C. Upon completion of the desired heating time, the transfer bag 112, now containing resealed erythrocytes, is reconnected, and the erythrocytes are pumped back into the separation bowl 106. Once in the bowl, the erythrocytes are washed with about 2 liters of physiological saline (supplied at about 150-200 ml/min) while the bowl is rotated at about 5600 rpm. Finally, the washed, loaded erythrocytes are transferred into the loaded erythrocyte bag 148. This is achieved by closing clamp 122 and opening clamp 134 while pumping the erythrocytes from the bowl at a rate of about 150 ml/min. The erythrocyte suspension is now ready for gravity infusion into a patient. As noted above, the patient may be either the patient who supplied the processed blood or another.

In the case in which drug-loaded erythrocytes are to be processed for the induction of their rapid recognition by macrophages, prior to pumping the loaded erythrocytes into bag 148, they are subjected to an additional washing step with a physiological saline solution containing 1mM ZnCl₂. Once contacted with ZnCl₂, the erythrocytes are transferred to the transfer bag 112 where they are contacted with BS³ (1-1.5 mM final concentration). After about 15 min, the erythrocytes are again washed in the rotating separation element 106 using approximately 1 liter of physiological saline. They are then transferred to the loaded erythrocyte bag 148, from where they may be infused into a patient.

If the loaded erythrocytes are to be stored longer than about 12-24 hours prior to infusion, the final collecting bag should contain a preservative. One preferred preservative is the CPD solution described above. Thus, approximately 10 ml of CPD solution may be provided in the final collecting bag.

The embodiment of FIGS. 1 and 2 is particularly preferred for the preparation of autologous carrier RBCs from a small volume (less than 100 ml) of blood from an individual patient at room temperatures, i.e., without requiring any heating or cooling of whole blood or any of the cellular or molecular components of blood. As used herein, "an individual patient" is not limited to humans and refers to any mammal to which carrier RBCs are

desired to be administered. Furthermore, as used herein, "autologous carrier RBCs" refers to erythrocytes isolated from a given individual patient that have been modified in vitro to encapsulate one or more biologically active agent(s) and which are intended to be re-introduced into the individual patient from which the erythrocytes were originally isolated. There are many advantages to the use of autologous carrier RBCs to treat individual patients and, in particular, undesirabl immune responses that might, for example, shorten the in vivo lifetime of the carrier RBCs are avoided by th use of erythrocytes derived from the individual patient.

Due to the higher concentration of added active agents in the carrier erythrocytes made by the method of the present invention, the carrier RBCs are not as rapidly depleted of such agents as those made by previously known methods. Moreover, due to the higher proportion of endogenous biological molecules that ar released from lysed RBCs but which, by the method of the present invention, are nevertheless re-introduced at a high concentration into the carrier RBCs, carrier erythrocytes made by the method of the present invention are not depleted for endogenous molecules needed for their long-term maintenance, as are carrier RBCs made by previously known methods. As a consequence, th carrier RBCs of the present invention possess an ability to persist and release active agents under physiological conditions for a longer period of time than previously described carrier RBCs.

The carrier RBCs of the present invention ar administered via injection into a mammal in order to provide the encapsulated biologically active agent(s) thereof to the mammal without further manipulation for an extended period of time. If systematic delivery is required, then the carrier RBCs are injected into the circulatory system of the mammal. Targeted delivery of the carrier RBCs is achieved by intramuscular injection in the region of interest.

In another embodiment of the invention, a "prodrug" (i.e., precursor to a biologically active agent) is incorporated into carrier RBCs that are then injected into a mammal. For some predrugs, endogenous activating agents present in the blood of the mammal will convert the predrug into a biologically active agent at a slow but steady rate. As an example, dexamethasone-21-phosphate (d-21P) may be encapsulated in carrier RBCs and, when the loaded RBCs are introduced into the circulatory system of a mammal, the d-21P is slowly converted into the chemotherapeutic drug dexamethasone. Because dexamethasone can cross the cell membrane of the carrier erythrocytes, (while d-21P cannot), this process assures that the mammal is provided with a constant level of the biologically active agent, (in this case dexamethasone), over some period of time. Alternatively, for predrugs for which no endogenous activating agents exist, an activating agent can be added to the blood of the mammal at a desired point in time. The activating agent enters the loaded erythrocytes and acti-

30

35

40

vates the predrug by, e.g., reacting with the prodrug to form a biologically active agent, or by catalyzing a ch mical reaction that converts th predrug. In a related embodiment, the activating agent may be encapsulated into erythrocytes which are introduced into a patient followed by the subsequent introduction of the predrug to the patient.

Carrier RBCs made by the method of the present invention are used to deliver a therapeutically effective amount of the biologically active agent(s) encapsulated therein to an individual patient. The term "therapeutically effective amount," for the purposes of the present invention, refers to the amount(s) of biologically active agent(s) effective to achieve an intended purpose. While individual needs vary, determination of optimal ranges for therapeutically effective amounts of such agents is within the skill of the art. Generally, the dosage required to provide a therapeutically effective amount of the composition can be determined by one of ordinary skill in the art and will vary depending on the age, health, physical condition, weight, extent of disease of the individual, frequency of treatment and the nature and scope of the desired effect. Where appropriate, a therapeutically effective amount can be delivered via multiple administrations of carrier RBCs over an appropriate period of time.

The carrier RBCs of the invention can be included in a pharmaceutical composition that also comprises additional substances, such as conventional excipients, i.e., pharmaceutically acceptable organic or inorganic carrier substances suitable for administration to a mammal via injection which do not deleteriously react with active compound. Suitable pharmaceutically acceptable carriers include, but are not limited to, water, salt solutions, polyethylene glycols, gelatin, etc. Additionally or alternatively, pharmaceutical preparations comprising the carrier RBCs of the invention can further comprise auxiliary agents, e.g., lubricants, preservatives, stabilizers, wetting agents, emulsifiers, salts for influencing osmotic pressure, buffers, and the like, which do not deleteriously react with the active compounds. Pharmaceutical compositions comprising the carrier RBCs of the invention may also optionally contain stabilizing agents.

One example of a polypeptide that is suitable for use with the present invention is ubiquitin, a polypeptide having a molecular weight of 8,600 Daltons. Ubiquitin is known to mark proteins for subsequent proteolytic degradation. Typically, the percentage of polypeptide that can be encapsulated into carrier erythrocytes (the encapsulation yield) is about 30%. In such instances, about 40% to about 60% of the starting erythrocytes were recovered.

One example of a predrug that is suitable for use with the present invention is d-21P. As with the polypeptide, the percentage of d-21P that can be encapsulated into carrier erythrocytes (the encapsulation yield) is about 30%. Again, about 40% to about 60% of the start-

ing erythrocytes were recovered.

In the case in which it is desired to obtain macrophage-targeted carrier RBCs, the following steps may be employed. First, the reconstituted, (i.e., loaded), RBCs are washed with saline containing 1 mM ZnCl₂. The ZnCl₂ acts as a clustering agent by inducing clustering of the transmembrane proteins of the RBCs. Next, a solution containing 1 mM bissulfosuccinimidisuberate (BS³) is added. BS³ acts as a cross-linking agent which causes protein clusters induced by ZnCl₂ to become irreversible even upon ZnCl₂ removal. Th incubation with BS³ is allowed to proceed for about 15 minutes. The resulting cross-linked erythrocytes are then washed with saline and concentrated by conventional methods.

Equivalents

Various modifications and alterations to this invention will become apparent to those skilled in the art without departing from the scope and spirit of this invention. It should be understood that this invention is not intended to be unduly limited by the illustrative embodiments and examples set forth herein and that such examples and embodiments are presented by way of example only with the scope of the invention intended to be limited only by the claims set forth herein as follows.

Where technical features mentioned in any claim are followed by reference signs, those reference signs have been included for the sole purpose of increasing the intelligibility of the claims and accordingly, such reference signs do not have any limiting effect on the interpretation of each element identified by way of example by such reference signs.

Claims

- A method for encapsulating at least one biologically active agent within a erythrocyte sample comprising the steps of:
 - a) providing a volume of erythrocytes;
 - b) at least partially lysing the erythrocytes;
 - c) concentrating the at least partially lysed erythrocytes;
 - d) contacting the concentrated, at least partially lysed erythrocytes with one or more biologically active agents; and
 - e) sealing the at least partially lysed erythrocytes to thereby encapsulate the biologically active agents within at least a portion of the erythrocytes.
- 2. A method as in claim 1, wherein the erythrocytes are swollen prior to the lysing step.
- 3. A method as in claim 1, wherein at least some of the erythrocytes are fully lysed.

25

30

45

- 4. A method for encapsulating at least one biologically active agent within a erythrocyte sample comprising the steps of:
 - a) exposing the erythrocytes to a first hypotonic solution to cause the erythrocytes to become swollen:
 - b) exposing the erythrocytes to a second hypotonic solution to cause the erythrocytes to become at least partially lysed;
 - c) concentrating the erythrocytes;
 - d) contacting the concentrated, at least partially lysed erythrocytes with one or more biologically active agents; and
 - e) exposing the at least partially lysed erythrocytes to a sealing solution to thereby encapsulate at least some of the biologically active agents within at least a portion of the erythrocytes.
- A method as in claim 4 wherein the first hypotonic solution comprises about 6 volumes of a physiological saline and about 4.5 volumes of distilled, sterile H₂O.
- A method as in claim 4 wherein the second hypotonic solution comprises about 6 volumes of physiological saline and about 8 volumes of distilled, sterile H₂O.
- A method as in claim 4 wherein the erythrocytes are concentrated using a hemofilter.
- 8. A method as in claim 4 wherein the one or more biologically active agents are selected from the group consisting of peptides, oligopeptides, polypeptides, proteins, hormones, corticosteroids, glucocorticoids, non-steroidal anti-inflammatory agents, glutathione, cytokines, toxins, oligonucleotides and other nucleic acids, and nucleoside analogs.
- A method as in claim 4 wherein the one or more biologically active agents are selected from the group consisting of dexamethasone-21-phosphate, prednisolone-21-phosphate, 6-mercaptopurine, azidothymidine, dideoxycytosine, and dideoxyinosine
- A method as in claim 4 wherein the sealing solution comprises a Phosphate-Inosine-Glucose-Pyruvate-Adenine solution.
- 11. A method as in claim 10 wherein about 5 ml of sealing solution is used for each aliquot of about 70-80 ml of at least partially lysed erythrocytes.
- 12. A method as in claim 11 wherein, while contacted

- with the sealing solution, the at least partially lysed erythrocytes are incubated at about 37 °C for about 30 minutes.
- 13. A method as in claim 4, wherein at least some of the erythrocytes are fully lysed when contacted with the second hypotonic solution.
- 14. A method for encapsulating at least one biologically active agent within a erythrocyte sample comprising the steps of:
 - a) providing a sample of erythrocytes from a donor;
 - b) suspending the erythrocytes in a first hypotonic solution to cause the erythrocytes to become swollen, the erythrocytes being present in the first hypotonic solution at a concentration of about 0.04 to about 0.05 ml erythrocytes per ml solution,
 - c) suspending the erythrocytes to a second hypotonic solution having an osmolality that is about 30 to about 60 mOsm less than that of the first solution, to cause the erythrocytes to become at least partially lysed;
 - d) concentrating the resulting suspension s that the resulting volume of erythrocytes is about 0.01 to about 0.067 times that of the volume of the first hypotonic solution;
 - e) contacting the concentrated, at least partially lysed, erythrocytes with one or more biologically active agents; and
 - f) exposing the at least partially lysed erythrocytes to a sealing solution to thereby encapsulate at least some of the biologically active agents within at least a portion of the erythrocytes, the sealing solution comprising a concentrated saline solution having a concentration such that the osmolality of the resulting solution is brought to about 1.9 to about 2.1 times that of the first hypotonic solution.
 - 15. A method as in claim 14, wherein the erythrocyt suspension is concentrated using a hemofilter.
 - 16. A method as in claim 14, wherein the one or more biologically active agents are selected from the group consisting of peptides, oligopeptides, polypeptides, proteins, hormones, corticosteroids, glucocorticoids, non-steroidal anti-inflammatory agents, glutathione, cytokines, toxins, oligonucleotides and other nucleic acids, and nucleoside analogs.
 - 17. A method as in claim 14, wherein the one or more biologically active agents are selected from the group consisting of dexamethasone-21-phosphate,

15

prednisolone-21-phosphate, 6-mercaptopurine, azidothymidine, dideoxycytosine, and dideoxyinosine.

18. A method as in claim 14, which further comprises 5 the step of treating the erythrocytes encapsulating at least one biologically active agent with a targeting agent such that the treated erythrocytes are preferentially directed to a specific tissue or cell type.

19. A method as in claim 18, wherein the targeting agent is ZnCl2 and bissulfosuccinimidilsuberate, and the treated erythrocytes are preferentially directed to macrophages.

- 20. Carrier erythrocytes made by the method of any one of claims 1-19.
- 21. A pharmaceutical composition containing erythrocytes encapsulating at least one biologically active agent, made by the method of any one of claims 1-
- 22. An apparatus for carrying out the method of any 25 one of claims 1-19.
- 23. An apparatus as in claim 22, wherein concentration of the at least partially lysed erythrocytes is achieved using a hemofilter.

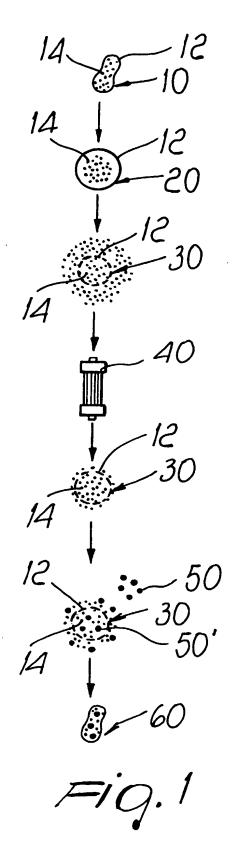
35

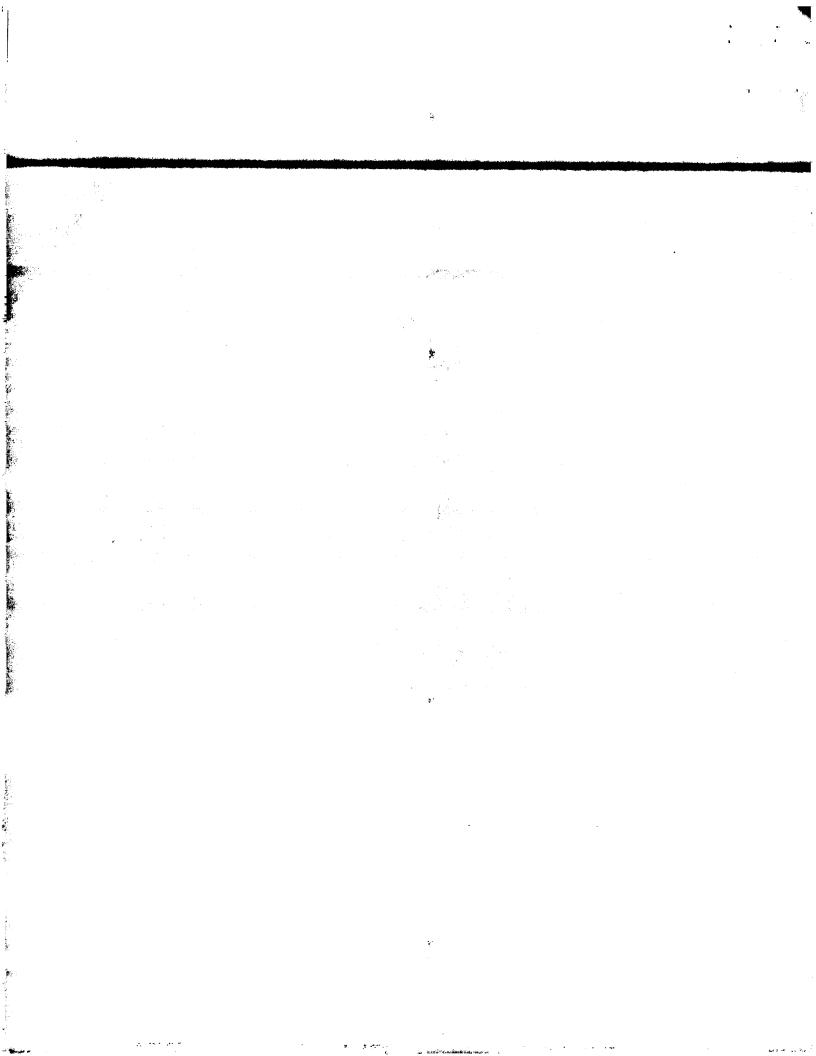
30

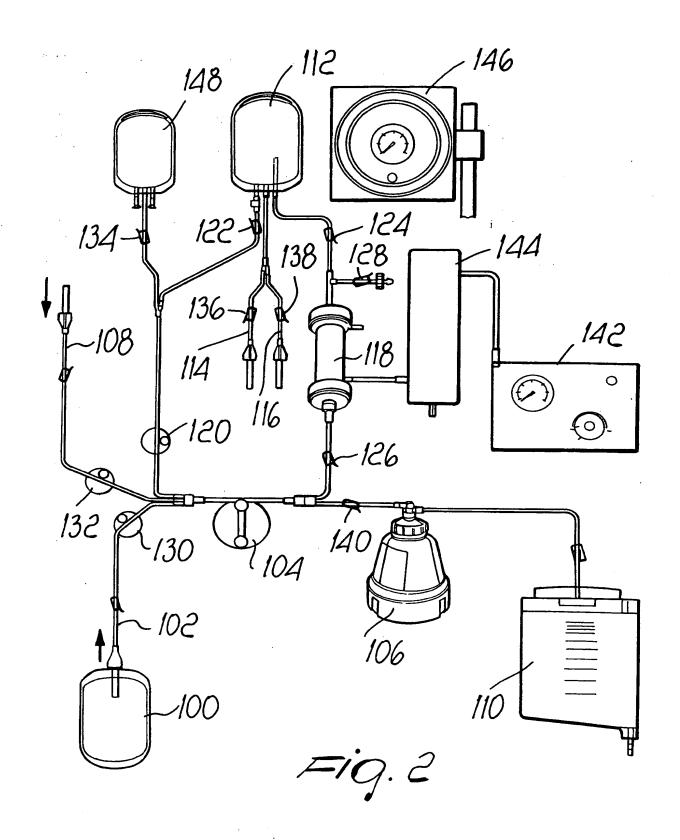
40

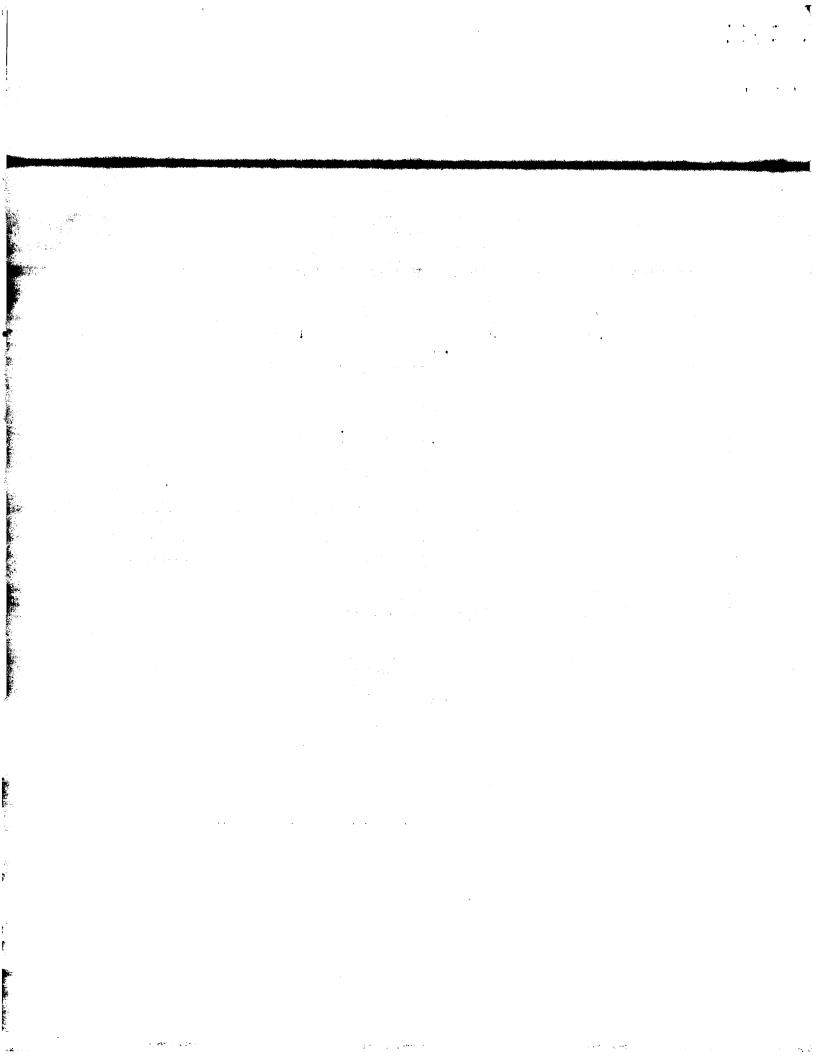
45

50











EUROPEAN SEARCH REPORT

Application Number EP 97 83 0211

May 1982 * column 2, line 13 - line 27 * * claim 1 * X FR 2 529 463 A (CENTRE NAT RECH SCIENT) 6 January 1984 * page 3, line 16 - line 25 * * page 6, line 1 - page 7, line 4 * * page 11, line 5 - line 14 * * page 24 - page 25; example 5 * X NARESH TALWAR ET AL: "ERYTHROCYTES AS CARRIERS OF PRIMAQUINE-PREPARATION: CHARACTERIZATION AND EVALUATION" JOURNAL OF CONTROLLED RELEASE, vol. 20, no. 2, 1 June 1992, pages 133-141, XP000269506 * page 134, column 2, paragraph 2 - page 135, column 1, paragraph 1 * TECHNICA	CLASSIFICATION APPLICATION	Relevant to claim					Category	
January 1984 * page 3, line 16 - line 25 * * page 6, line 1 - page 7, line 4 * * page 11, line 5 - line 14 * * page 24 - page 25; example 5 * X NARESH TALWAR ET AL: "ERYTHROCYTES AS CARRIERS OF PRIMAQUINE-PREPARATION: CHARACTERIZATION AND EVALUATION" JOURNAL OF CONTROLLED RELEASE, vol. 20, no. 2, 1 June 1992, pages 133-141, XP000269506 * page 134, column 2, paragraph 2 - page 135, column 1, paragraph 1 * X JAIN S K ET AL: "MAGNETICALLY RESPONSIVE DICLOFENAC SODIUM-LOADED ERYTHROCYTES: PREPARATION AND IN VITRO CHARACTERIZATION" JOURNAL OF MICROENCAPSULATION, vol. 11, no. 2, 1 March 1994, pages 141-151, XP000425614 * page 143 * EP 0 517 986 A (EUROP COMMUNITIES) 16 December 1992 * page 7, line 17 - line 21 * FR 2 678 512 A (NOVACELL) 8 January 1993 * page 1, line 1 - line 3 * * page 8, line 6 - line 17 *	 1-3, A61K9/50		·	May 1982 * column 2, line 13 - lin	X			
CARRIERS OF PRIMAQUINE-PREPARATION: CHARACTERIZATION AND EVALUATION" JOURNAL OF CONTROLLED RELEASE, vol. 20, no. 2, 1 June 1992, pages 133-141, XP000269506 * page 134, column 2, paragraph 2 - page 135, column 1, paragraph 1 * X JAIN S K ET AL: "MAGNETICALLY RESPONSIVE DICLOFENAC SODIUM-LOADED ERYTHROCYTES: PREPARATION AND IN VITRO CHARACTERIZATION" JOURNAL OF MICROENCAPSULATION, vol. 11, no. 2, 1 March 1994, pages 141-151, XP000425614 * page 143 * EP 0 517 986 A (EUROP COMMUNITIES) 16 December 1992 * page 7, line 17 - line 21 * FR 2 678 512 A (NOVACELL) 8 January 1993 * page 1, line 1 - line 3 * * page 8, line 6 - line 17 *	·		e 4 *	January 1984 * page 3, line 16 - line * page 6, line 1 - page 7 * page 11, line 5 - line				
JAIN S K ET AL: "MAGNETICALLY RESPONSIVE DICLOFENAC SODIUM-LOADED ERYTHROCYTES: PREPARATION AND IN VITRO CHARACTERIZATION" JOURNAL OF MICROENCAPSULATION, vol. 11, no. 2, 1 March 1994, pages 141-151, XP000425614 * page 143 * EP 0 517 986 A (EUROP COMMUNITIES) 16 December 1992 * page 7, line 17 - line 21 * FR 2 678 512 A (NOVACELL) 8 January 1993 * page 1, line 1 - line 3 * * page 8, line 6 - line 17 *			TION: N"	CARRIERS OF PRIMAQUINE-PR CHARACTERIZATION AND EVAL JOURNAL OF CONTROLLED REL vol. 20, no. 2, 1 June 19 pages 133-141, XP00026950 * page 134, column 2, par				
DICLOFENAC SODIUM-LOADED ERYTHROCYTES: PREPARATION AND IN VITRO CHARACTERIZATION" JOURNAL OF MICROENCAPSULATION, vol. 11, no. 2, 1 March 1994, pages 141-151, XP000425614 * page 143 * EP 0 517 986 A (EUROP COMMUNITIES) 16 December 1992 * page 7, line 17 - line 21 * FR 2 678 512 A (NOVACELL) 8 January 1993 * page 1, line 1 - line 3 * * page 8, line 6 - line 17 *	TECHNICAL FIE SEARCHED	1 2 20	DECDONCINC					
December 1992 * page 7, line 17 - line 21 * FR 2 678 512 A (NOVACELL) 8 January 1993 * page 1, line 1 - line 3 * * page 8, line 6 - line 17 *	A61K		ROCYTES:	DICLOFENAC SODIUM-LOADED PREPARATION AND IN VITRO JOURNAL OF MICROENCAPSULA vol. 11, no. 2, 1 March 19 pages 141-151, XP00042561				
* page 1, line 1 - line 3 * * page 8, line 6 - line 17 *		19	IES) 16	December 1992	[1			
-/		1-23		* page 1, line 1 - line 3	* page 1, line 1 -			
			-/					
The present search report has been drawn up for all claims			il claims	The present search report has been drawn				
Place of search Date of completion of the search Examiner	 Examiner		mpletion of the search	, , === = = = = = = = = = = = = = = = =				
THE HAGUE 20 October 1997 Boulois, D	lois, D	Bou	ctober 1997	THE HAGUE				
CATEGORY OF CITED DOCUMENTS X: particularly relevant if taken alone Y: particularly relevant if controlled with another document of the same category A: technological background O: non-written disclosure A: member of the same patent family, corresponding	 hed on, or	ment, but publis the application other reasons	E : earlier patent door after the filing date D : document cited in L : document cited for	cularly relevant if taken alone cularly relevant if combined with another ment of the same category cological background	X : partice Y : partice dooun A : techni			



EUROPEAN SEARCH REPORT

Application Number EP 97 83 0211

	DOCUMENTS CONSID	ERED TO BE RELEVANT			, a - (
Category	Citation of document with it of relevant pass	ndication, where appropriate,	Relevant to claim	CLASSIFICAT APPLICATION	TON OF THE
X	ROPARS C. ET AL: "	Resealed Red Blood od Transfusion Product* L, 43887 h 7 - page 84,	1-3,20, 21	# ·	
x	RECHSTEINER M.C.: Red Cells" EXPERIMENTAL CELL R vol. 93, 1975, pages 487-492, XP00 * page 488, column	2043888	1-6,8, 20,21		当 (*) (*) (*) (*) (*)
	MAGNANI M. ET AL: Dexamethasone to Ma DRUG DELIVERY, vol. 2, no. 3/4, 19 pages 151-155, XP00 * page 151, column	crophages	9	TECHNICAL SEARCHED	FIELDS (int.Cl.8)
	Clustering" MOL. CEL. BIOCHEM., vol. 144, no. 1, 199 pages 53-59, XP0020	by Membrane Protein 95, 43890 , paragraph 3 - page raph 1 *	18,19		
	The present search report has b	-/			
	Place of search	Date of completion of the search		Examiner	· ·
	THE HAGUE	20 October 1997	Bou	ois, D	
CAT X : partic Y : partic docum A : techn	TEGORY OF CITED DOCUMENTS was in the same of the same was in the same one of the same to logical background written disclosure	T: theory or principle E: earlier patent docu after the filing date E: D: document cited in L: document dead for	underlying the Imment, but publish the application other reasons.	rention led on, or	· · ·

12

EPO FORM 1503 03.82 (P04C01)



EUROPEAN SEARCH REPORT

Application Number EP 97 83 0211

Category	DOCUMENTS CONSIDER Citation of document with inci-	cation, where appropriate,	Relevant	CLASSIFICATION OF THE
- aayury	of relevant passage	98	to claim	APPLICATION (Int.CI.5)
X	MAFNANI M. ET AL: "antiretroviral nucleo phosphorylated form to vitro and in vivo stu PROC. INT. ACAD. SCI. vol. 89, July 1992, pages 6477-6481, XPOG * page 6477, column 2 6488, column 1, parag	oside analogues in to macrophages : In udies" . USA, 02043972 2, paragraph 4 - page	1-4,8, 10,20,21	ı
				TECHNICAL FIELDS SEARCHED (Int.Ct.6)
				*
		·		
		• .		
	The second of		1	
	The present search report has been	Date of completion of the search		Exeminer
•	THE HAGUE	20 October 1997	Boul	ois, D
X : partio Y : partic docum	TEGORY FOITED DOCUMENTS ularly relevant if taken alone ularly relevant if combined with another nent of the same category placing leadmounts.	T: theory or principl E: earlier patent do after the filing dat D: document cited i L: document cited i	e underlying the involument, but publish to not application or other reasons	rention ed on, or
A : technological background C : non-written disclosure P : intermediate document		& : member of the s	ame patent family, o	

in the second se

.

.